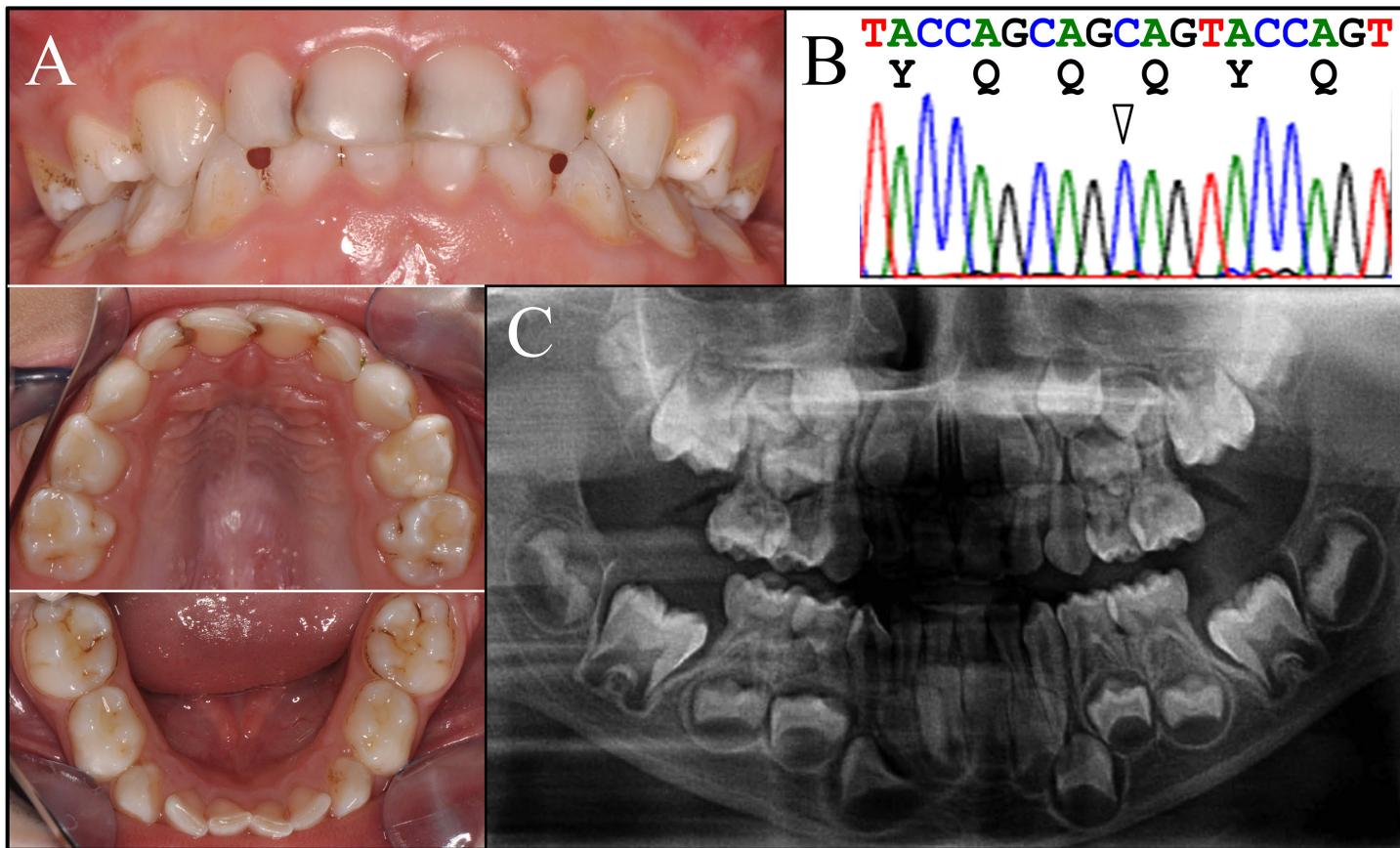


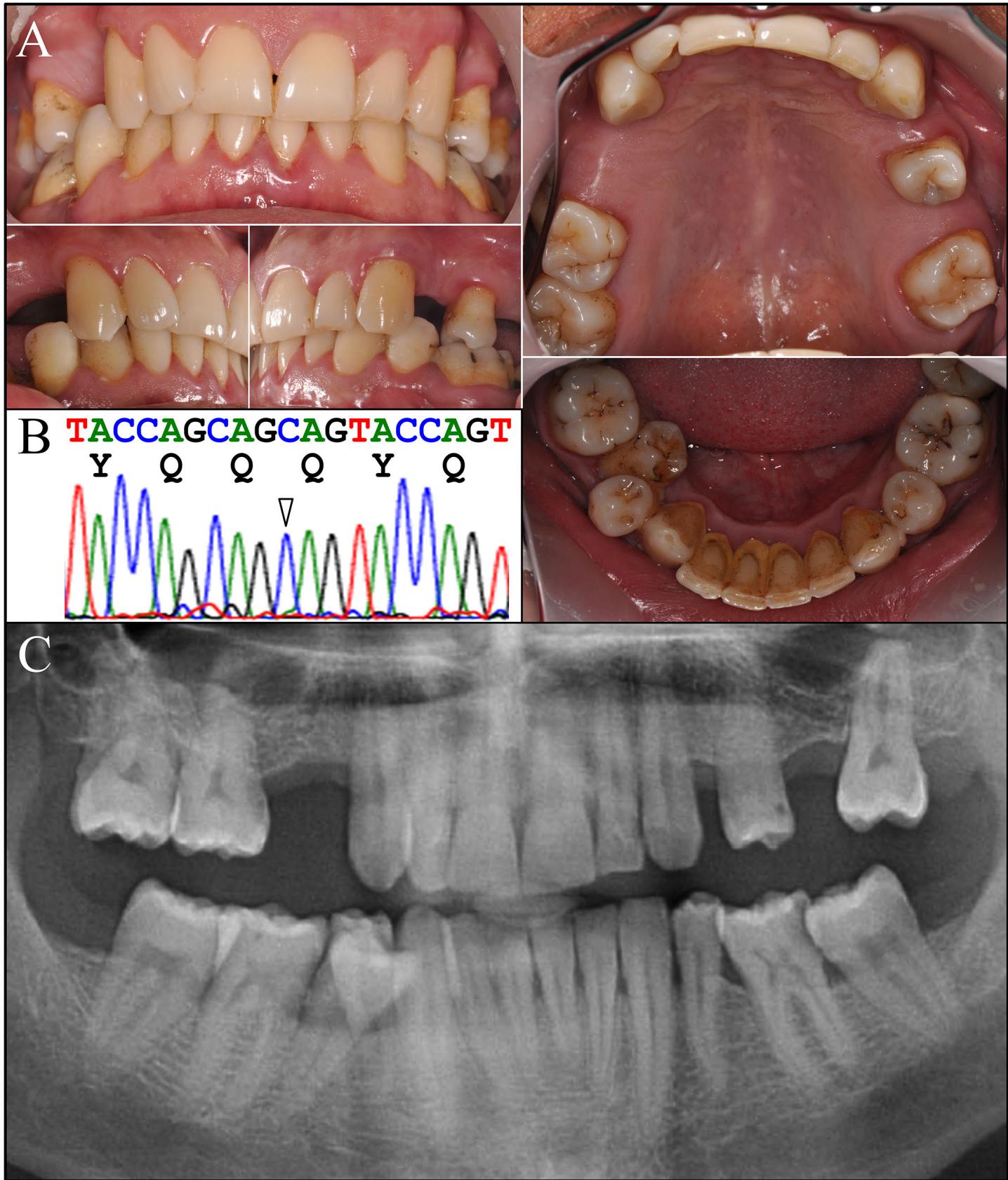
**Fig. S1.** *FAM83H* disease-causing mutations. *FAM83H* gene structure: numbered boxes indicate exons; introns are lines connecting the exons. The numbers above each intron indicate the length of the intron in basepairs (bp). The numbers below each exon show the length of the exon in bp and below that the range of amino acids encoded by it. Shaded exon regions are non-coding. The 20 reported *FAM83H* nonsense or frameshift mutations are located between the sites marked 1 and 20 in bold. The gene numbers start from the first nucleotide of the National Center for Biotechnology Information (NCBI) *FAM83H* genomic reference sequence NG\_016652.1. The cDNA numbers start from the translation initiation site of *FAM83H* cDNA reference sequence NM\_198488.3.

## References (Figure S1)

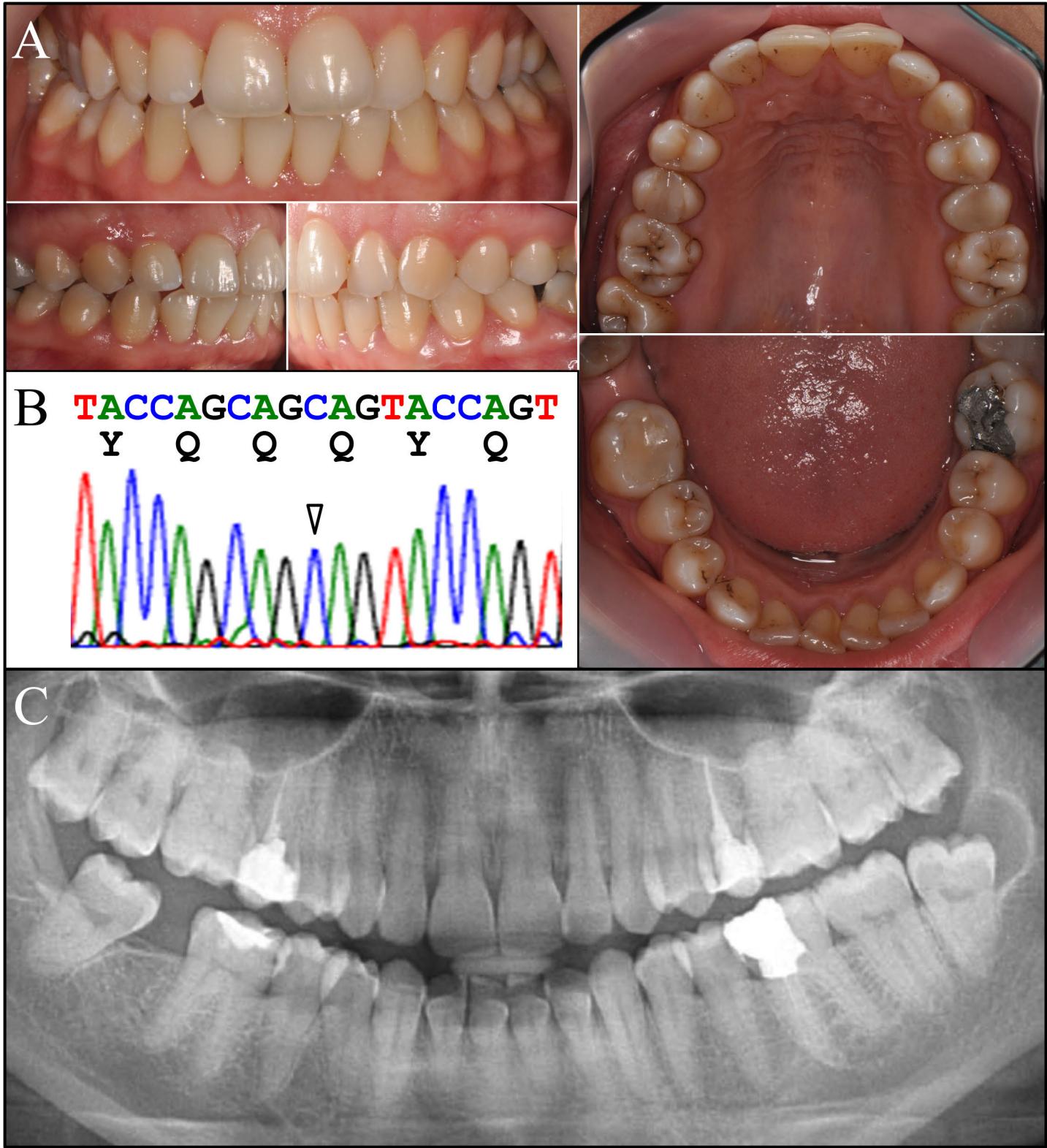
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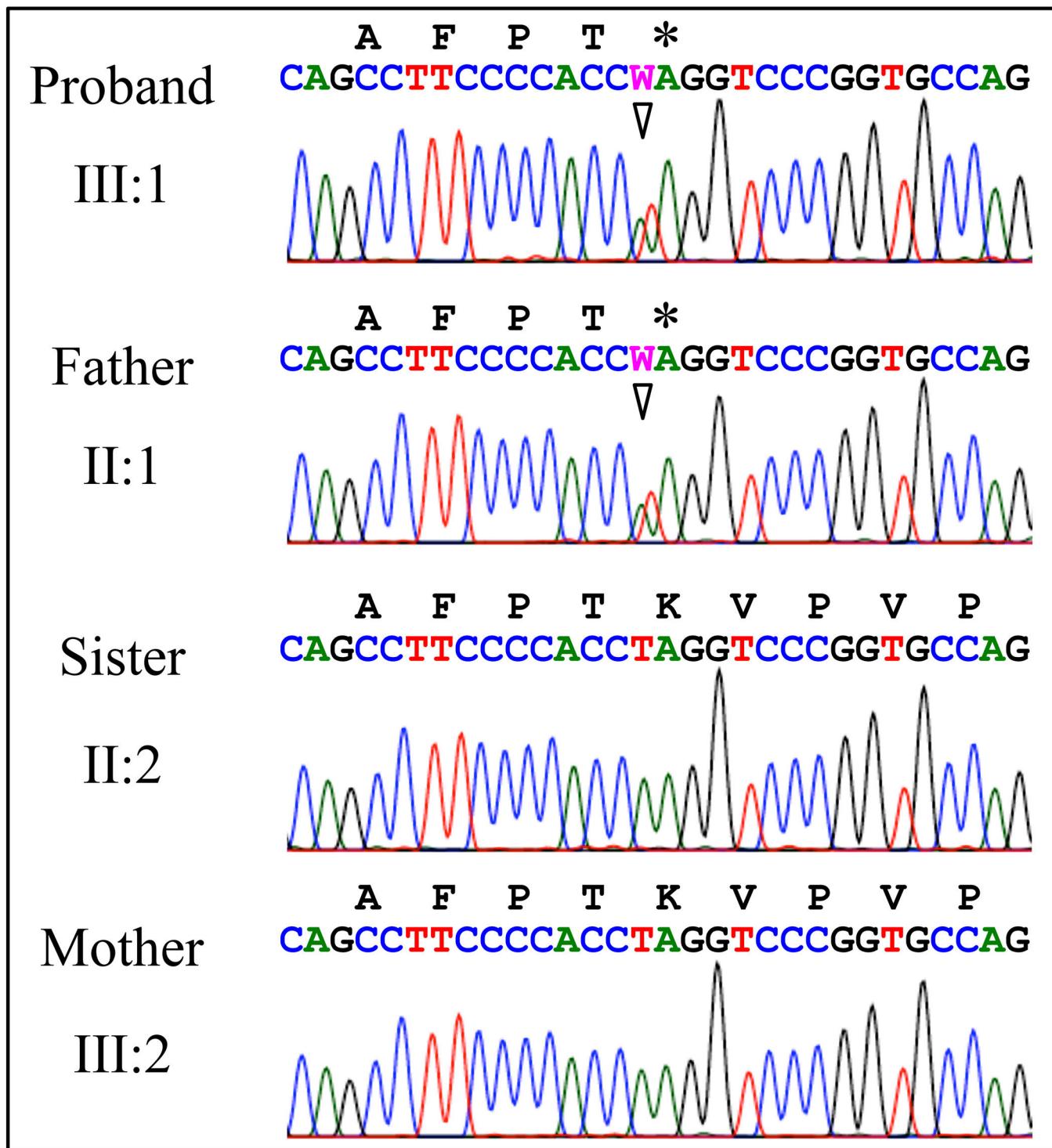
**Fig. S2.** Family 1. Unaffected Brother (II:2). **A:** Oral photographs. **B:** Chromatogram showing normal *FAM83H* sequence in both alleles at the site (c.1369C>T) mutated in the proband. **C:** Panorex.



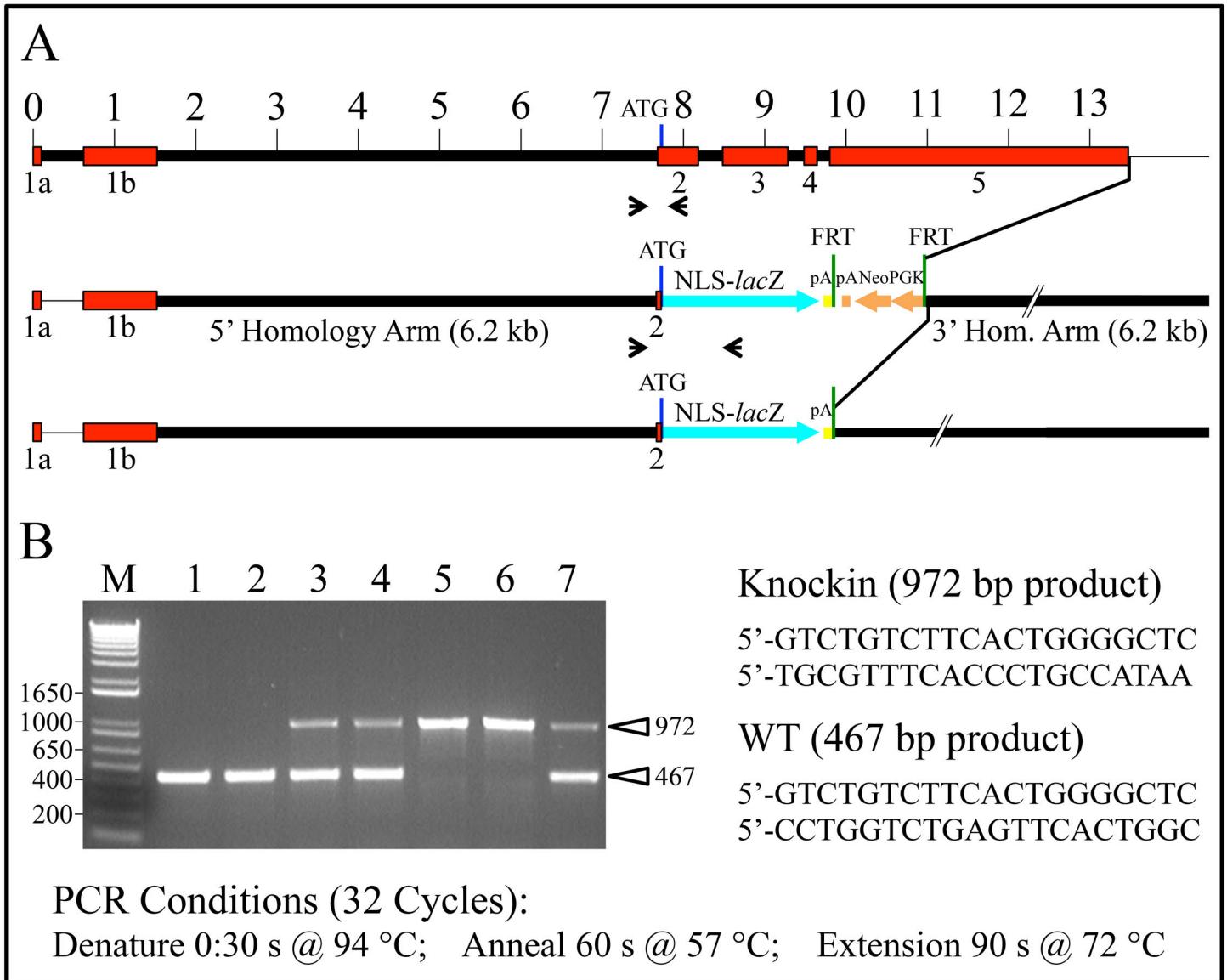
**Fig. S3.** Family 1. Unaffected Father (I:1). **A:** Oral photographs. **B:** Chromatogram showing normal *FAM83H* sequence in both alleles at the site (c.1369C>T) mutated in the proband. **C:** Panorex.



**Fig. S4.** Family 1. Unaffected Mother (I:2). **A:** Oral photographs. **B:** Chromatogram showing normal *FAM83H* sequence in both alleles at the site (c.1369C>T) mutated in the proband. **C:** Panorex.



**Fig. S5.** Family 2 Chromatograms. The affected proband (III:1) and affected father (II:1) both showed the single allele *FAM83H* truncation mutation (g.11199A>T, c.1915A>T, p.Lys639\*) that was absent from unaffected mother (II:2) and unaffected sister (III:2) and therefore segregated with the disease phenotype. W = A or T.



**Fig. S6.** *Fam83h* Knockin Construct and Genotyping Strategy. **A:** *FAM83H* gene structure (top): The numbered boxes (red) are exons. The numbers 1-13 indicate thousands of bp. Zero on the left marks the 5' end of the mouse *Fam83h* genomic reference sequence NG\_016652.1. Exons 1a and 1b are expressed from alternative promoters and are not translated. Translation initiation begins in exon 2 (ATG). The thin line following exon 5 indicates downstream untranscribed sequence not included in the genomic reference sequence. The first *Fam83h* knockin structure (middle). The entire *Fam83h* coding region starting at the translation initiation codon and extending to the end of exon 5 was replaced by the mouse *lacZ* coding region that was modified by adding a nuclear localization signal (NLS) and two downstream polyadenylation signals (pA, yellow). The Neomycin (*Neo*) and selection Protein kinase C (*PKC*) selection genes and associated downstream sequences (pA, brown) were bracketed by flippase recognition target (FRT) sites. Arrowheads indicate the PCR primer annealing sites used for genotyping. Final *Fam83h* knockin structure (bottom). Mating with flippase (*FLP*) deleter mice resulted in site-directed recombination and germline deletion of the *Neo* and *PKC* selection marker genes. **B:** 1% agarose gel stained with ethidium bromide showing typical genotyping results. Lanes M, molecular weight marker; lanes 1-2, wild-type; lanes 3-4, 7 heterozygotes; lanes 5-6, homozygous knockin mice. The same 5' primer (that annealed near the 5' end of intron1) was mixed with two reverse primers (one specific for the wt, the other for the knockin) for genotyping. The next figure shows the exact sequences that were deleted and inserted.

A

Exon 1a (nucleotides 1-93) is non-coding.

ACCTGGAGCGACTCCTCGAAACAGGAGGCCCTGGGAAGGAAGCTCAGATGAGAGTTGGTCTTGACCCTCAGCTCCACCCCTGGGACTAACAG

Intron 1a (nucleotides 94 to 7774) includes Exon 1b and Intron 1b.

gtggggcacgtgagggtcccttaggcctgtttaatttaaagggggtgctggactcccaatcatggcagttagtcaggcccacacccatcccaggagccaagctagacttcctttatctagactttgaaggctcagtgttccaccgcacgggtgtggcccaagtgtctactgtctggcctaggggtggccatctctctggcaaaactctgtctggcagggttcgtccgggtaggcctgggtccacgtgttcagtggtcgatgggtggatgacactatggccagtgagggtctggcaggtaggtggctgtgcatttggatgcagctcttgtctgtggccagtttagttcccgccacagagactgagggtgggtaaaggggggggggctgtgtgcacaccacccaggcaggaccaggcaagtcctcaagcagcagtttagctactgagagccgcggcccccagttctcagtctccctcccaagtctggccagtgaagtctgtggctgtaaagtctcgcacccctgtgtcagcg

Exon 1b (nucleotides 632 to 1540) is non-coding.

tgttagtcagcctcccgctagggtgagccactgtgccccaagtctataaactcttgtgtggcccagtgcacagactgggactgtgggcataaggcagagc  
cagggccctggctgtaccgggtcgagatcagccaagggttagcacagccacttggacagcgggagcagagggtctgaacctggctgagagcaagcaaccttgc  
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aactcaaggccag

### Intron 1b (nucleotides 1541 to 7774).

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 ccgtgtgtgtccacttgtggcatqaqtaaqtgtggctactgcccctagtaatqcatttctgcacctccag

Exon 2 (nucleotides 7775 to 8236).

gcccccgtggccccaac

**atggcccgctcgcccagagcagctcgcaaaaaactggcacctgggtacctgccacct**  
M A R R S Q S S S Q G D N P L A P G Y L P P  
cactacaaagaatattaccgcctagcggtggatgcattgactgagggtggccagaagcctacaac  
H Y K E Y Y R L A V D A L T E G G P E A Y N  
cgcttcttggcatctgagggggcacctgacttcctgtgcctgaggaacttggAACACGTgagccgc  
R F L A S E G A P D F L C P E E L E H V S R  
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H L Q P P Q Y V A R E P P E G T P S D V D M  
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W P L T F G F Q G T E V T T L V Q P P P P D  
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S P S I K D E A R R M I R S A O O

Intron 2 (nucleotides 8237 to 9037)

### Exon 3 (nucleotides 9038 to 9202)

Exon 5 (nucleotides 5956 to 6101)  
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V V A V M D M F T D V D L L S E V L E A A  
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A R R V P V Y I L L D E M N A Q H F L D M A  
gacaagtgtcgcgtaaacctgcatcatgtggac  
D K C R V N L H H V D

### Intron 3 (nucleotides 9203 to 9271)

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### Exon 4 (nucleotide 9272 to 9396)

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F L R V R T V A G P T Y Y C R T G K S F K G  
catctaaaggagaagtcttgcgtgtggactgtgccgtagtgtatgagcggcagttatag  
H L K E K F L L V D C A V V M S G S Y S

#### Intron 4 (nucleotide 9397 to 9587)

```
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```

## Exon 5 (nucleotide 9588 to 13245)

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 D E E F R I L F A Q S E P L V P S A G A L A R M  
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S K K \*

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qattqqqqaqactqqqcttgcataataaaqaatcqggacacqcttctgtqa

3' Untranscribed sequence (nucleotides 13246 to 14337; not in gene reference sequence). This sequence and the "a" that precedes it follow the NLS-*lacZ* insert highlighted in cyan below.

**B**

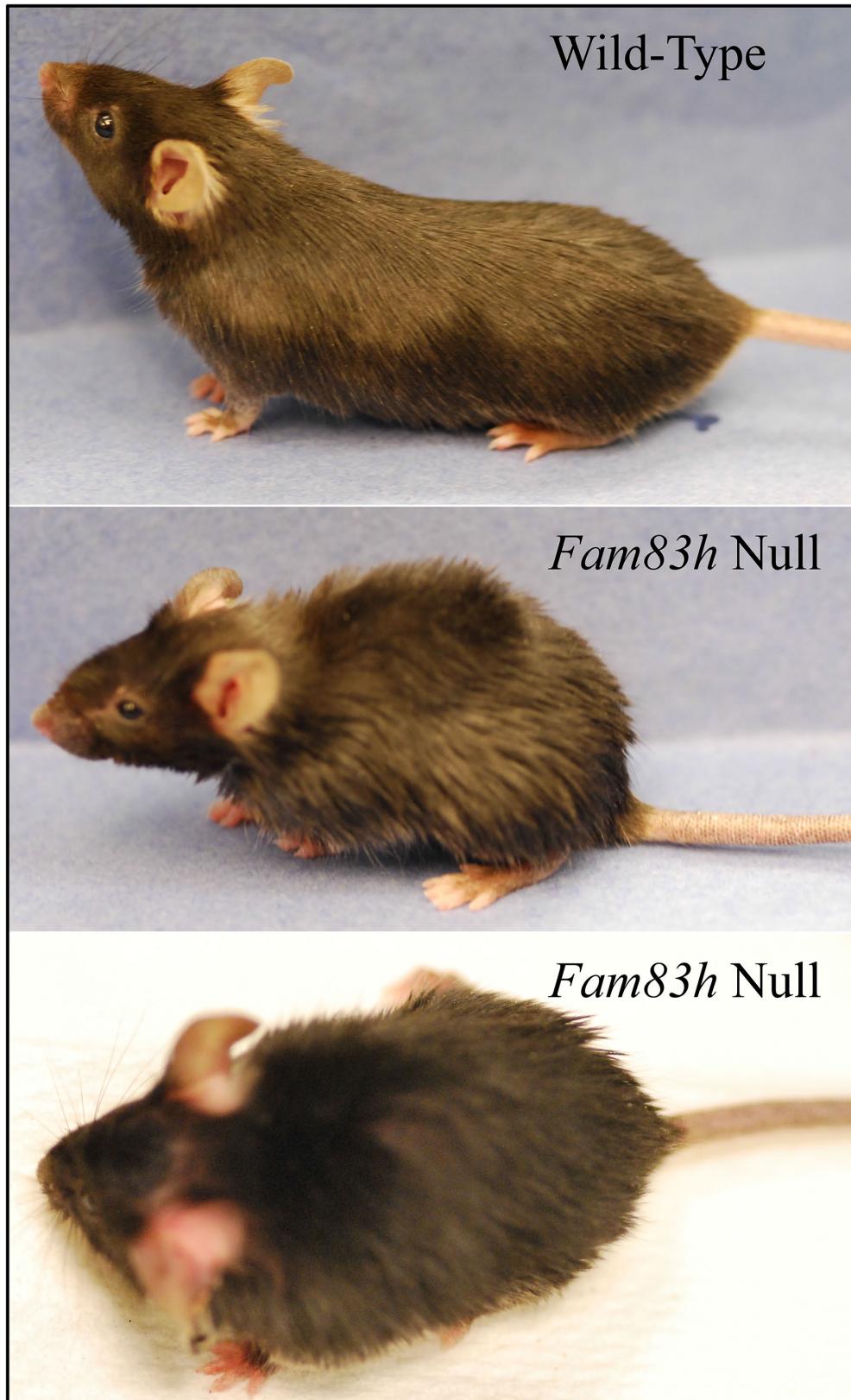
Area highlighted in yellow was replaced with the following NLS-*lacZ* sequence:

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 G V T Q L N R L A A H P P F A S W R N S E E A R  
 ACCGATGCCCTTCCCAACAGTTGCGCAGCCTGAATGGCGAATGGCGCTTGCCCTGGTTCCGGCACCAAGAA  
 T D R P S Q Q L R S L N G E W R F A W F P A P E  
 GCGGTGCCGGAAAGCTGGCTGGAGT GCGATCTCCTGAGGCCGATACTGTCGTGCTCCCTCAAACGGCAG  
 A V P E S W L E C D L P E A D T V V V P S N W Q  
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 M H G Y D A P I Y T N V T Y P I T V N P P F V P  
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 T E N P T G C Y S L T F N V D E S W L Q E G Q T  
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 R I I F D G V N S A F H L W C N G R W V G Y G Q  
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 D S R L P S E F D L S A F L R A G E N R L A V M  
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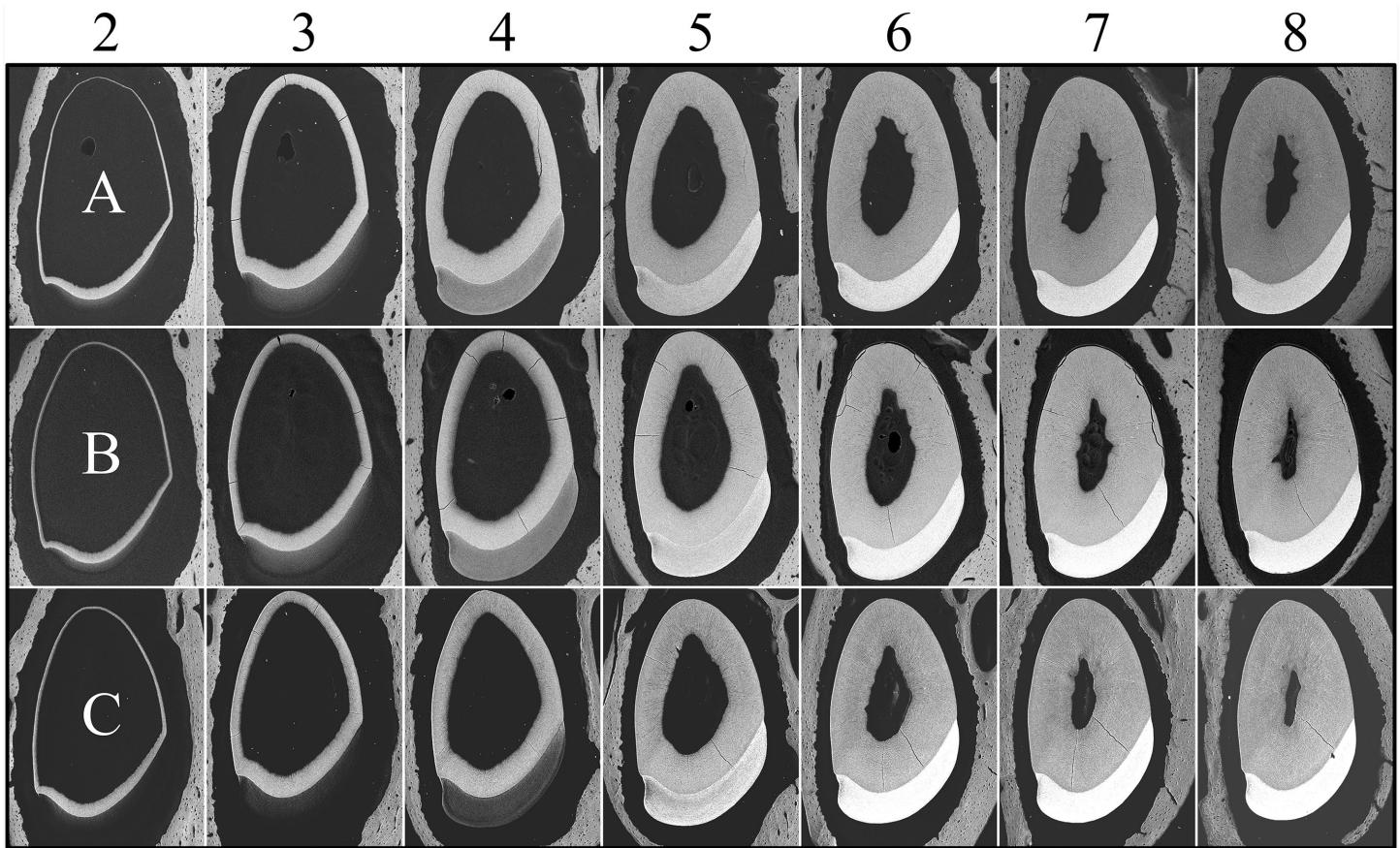
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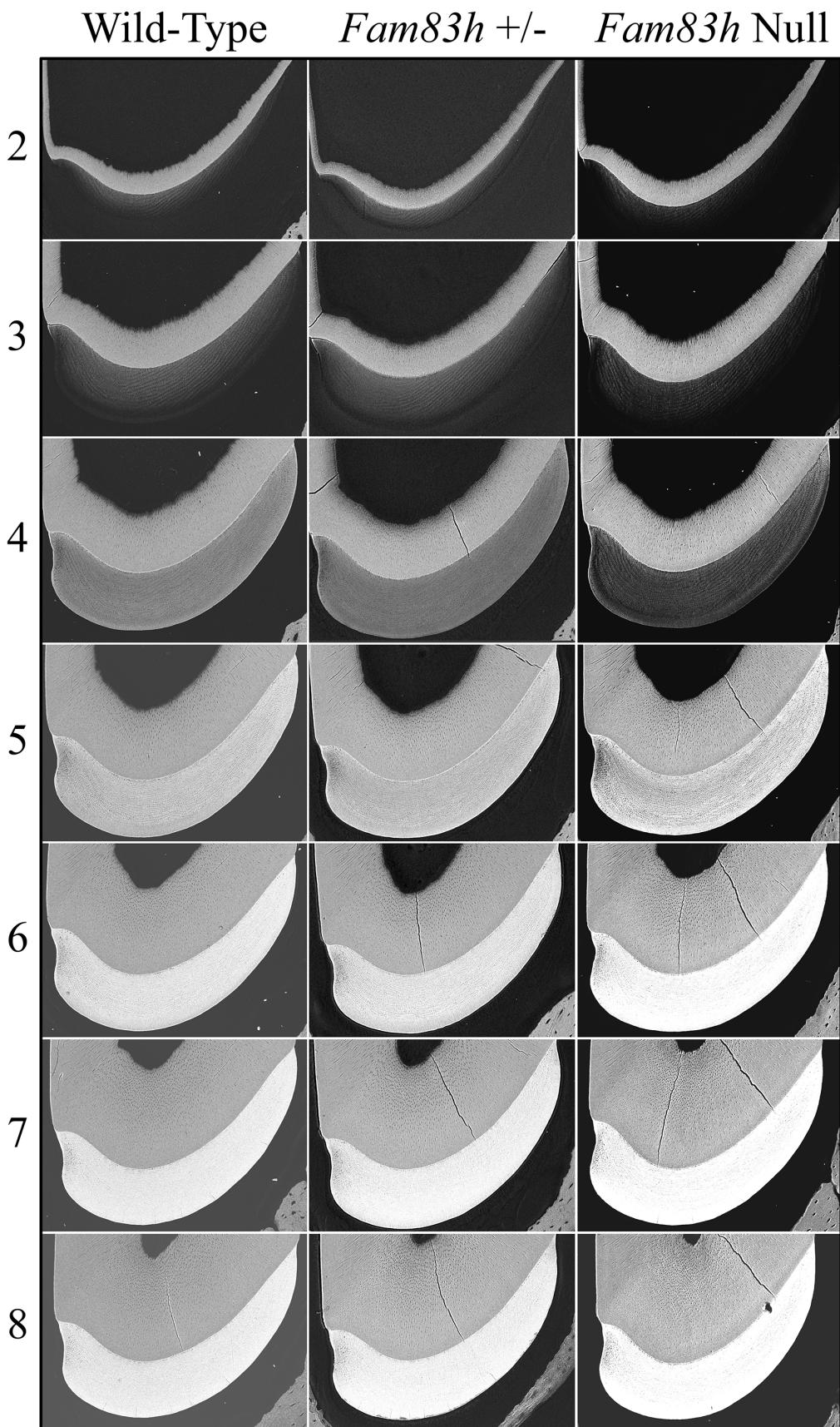
**Fig. S7.** Mouse *Fam83h* Wild-Type and NLS-*lacZ* Knockin Sequences. **A:** The NCBI genomic reference sequence NC\_000081.6 for mouse strain C57BL/6J starts with the first nucleotide of exon 1a, which is found on the 5' end transcript variant 2 (TV2) and ends 1 bp after exon 5. The entire coding region and all but a single nucleotide of the 3' untranslated region (highlighted in yellow) was deleted. **B:** The NLS-*lacZ* coding sequence and 3' untranslated region that included two polyadenylation signals (AATAAA, underlined) and the FRT sequence (bold) that remained following flippase recombination. Thus the wild-type mice differ only by the replacement of the sequence in yellow (in **A**) with the sequence in blue (in **B**).



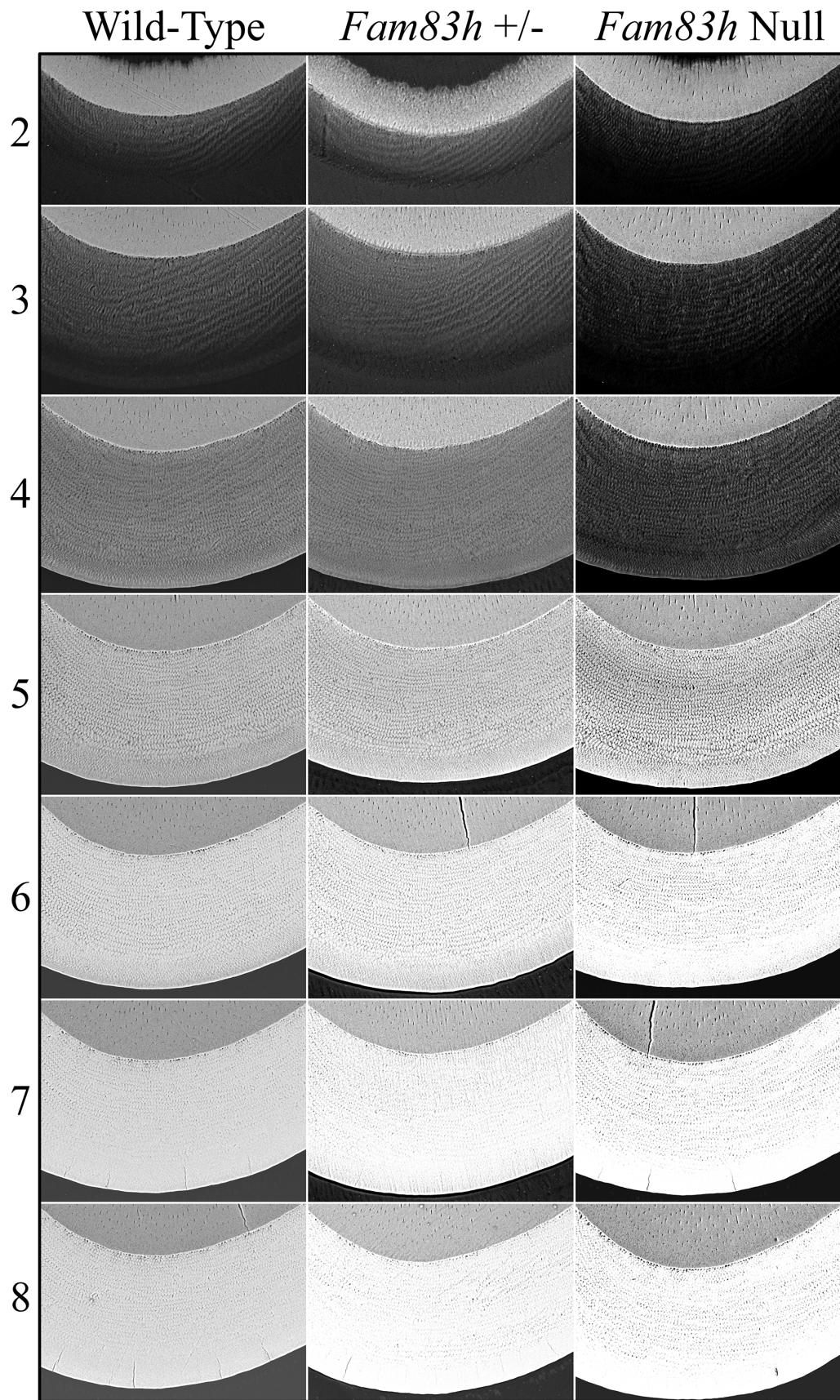
**Fig. S8.** Wild-type and *Fam83h* null mice at 7-weeks. The *Fam83h* null mice that survive to 7-weeks are smaller than the wild-type. Their fur tends to shed and coat hair gets caught up in the gingival crevice of the mandibular incisors.



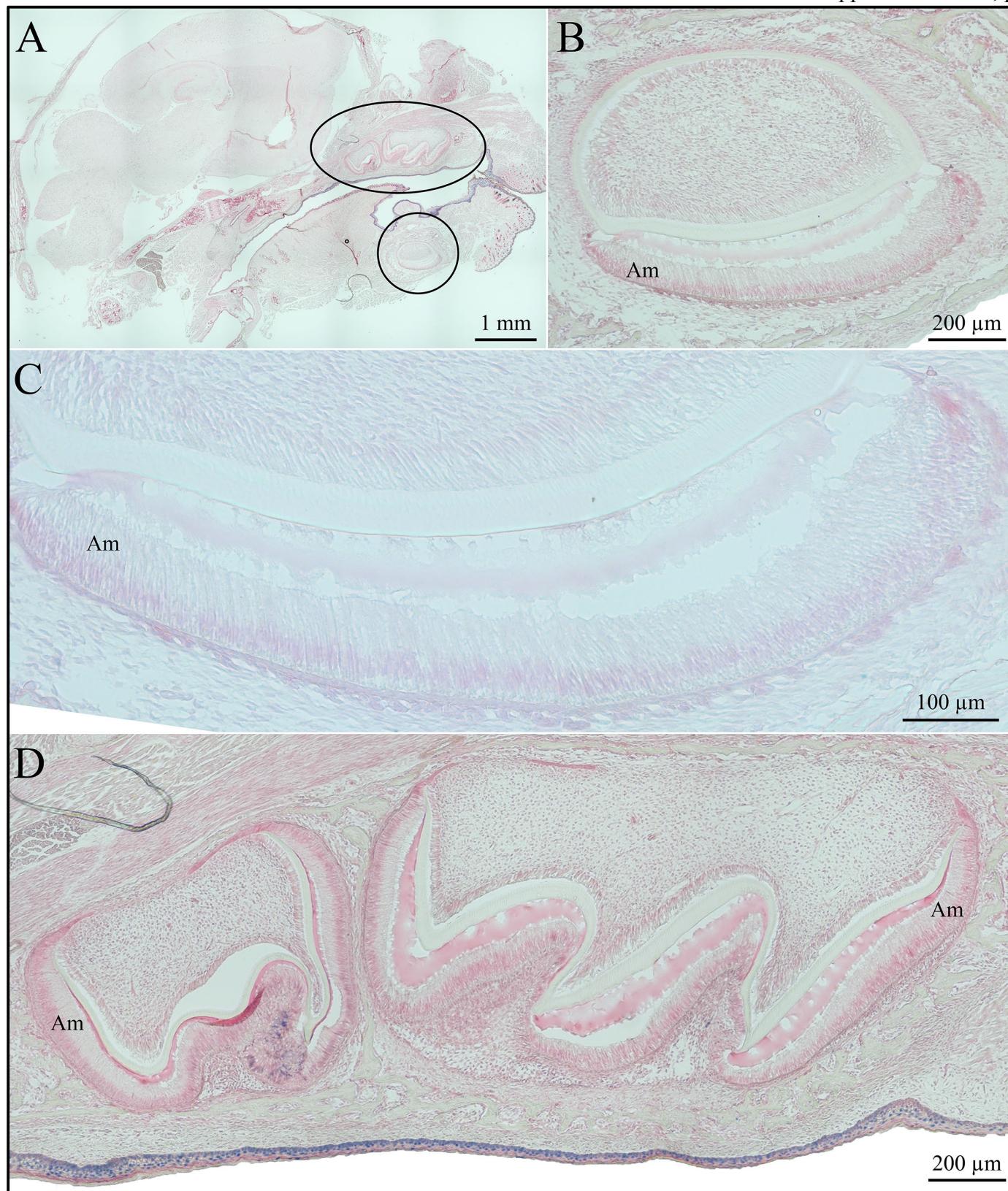
**Fig. S9.** bSEM Images of Mandibular Incisor Cross Sections at 7-weeks (lower magnification). Mandibular incisors display all stages of enamel formation. The incisors are cross-sectioned at 1 mm intervals and examined by bSEM. The basal ends are on the left. **A:** wild-type; **B:** *Fam83h<sup>+/−</sup>*; **C:** *Fam83h<sup>−/−</sup>*. Levels 2 and 3 show the secretory stage of amelogenesis when the enamel layer is expanding. Levels 4 through 8 show the maturation stage when the enamel layer no longer expands. During this stage the mineral ribbons deposited during the secretory stage grow in width and thickness and the enamel layer becomes increasingly mineralized. Level 8 is where the incisor reaches the level of the alveolar crest, still prior to eruption. The enamel layer appears to be fully mineralized in all 3 genotypes. The pulp space in the null mouse is smaller, suggesting that the incisors erupted more slowly than in the wild-type (giving the odontoblasts more time to add dentin).



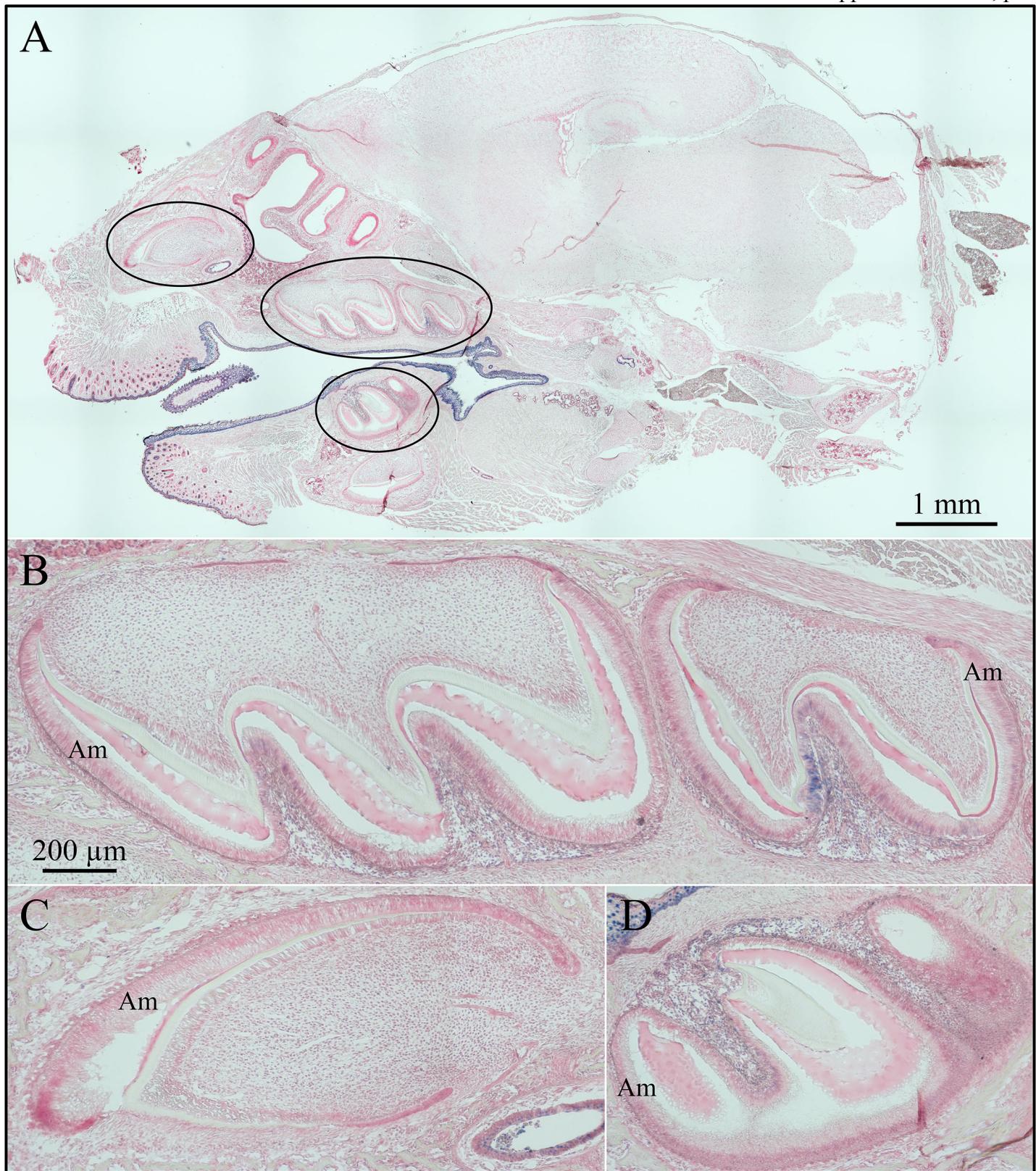
**Fig. S10.** bSEM Images of Manibular Incisor Cross Sections at 7-weeks (higher magnification). The enamel layer appears to be fully mineralized in all 3 genotypes.



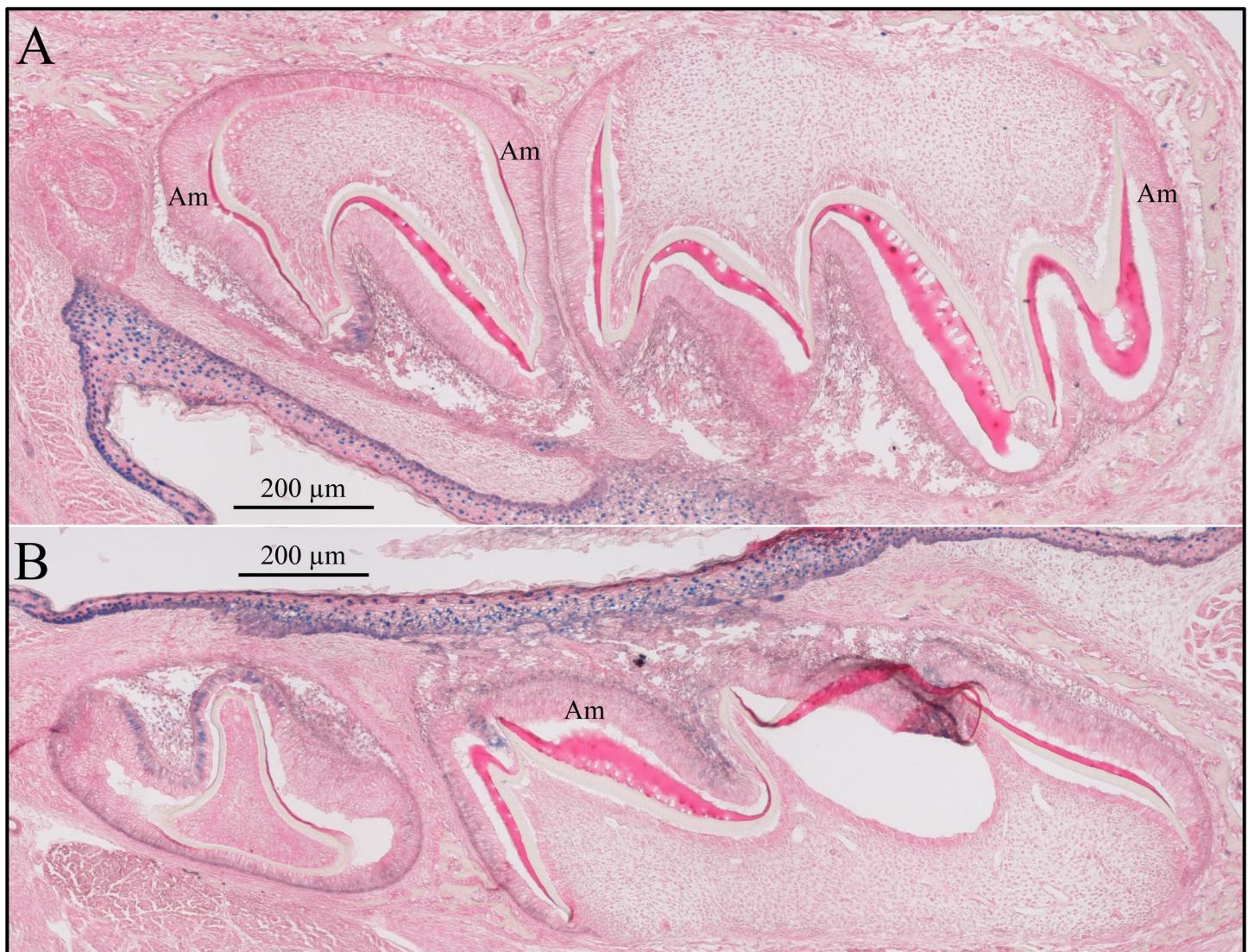
**Fig. S11.** bSEM Images of Manibular Incisor Cross Sections at 7-weeks (highest magnification). The enamel layer appears to be fully mineralized in all 3 genotypes. The enamel thickness is the same, suggesting normal development during the secretory stage.



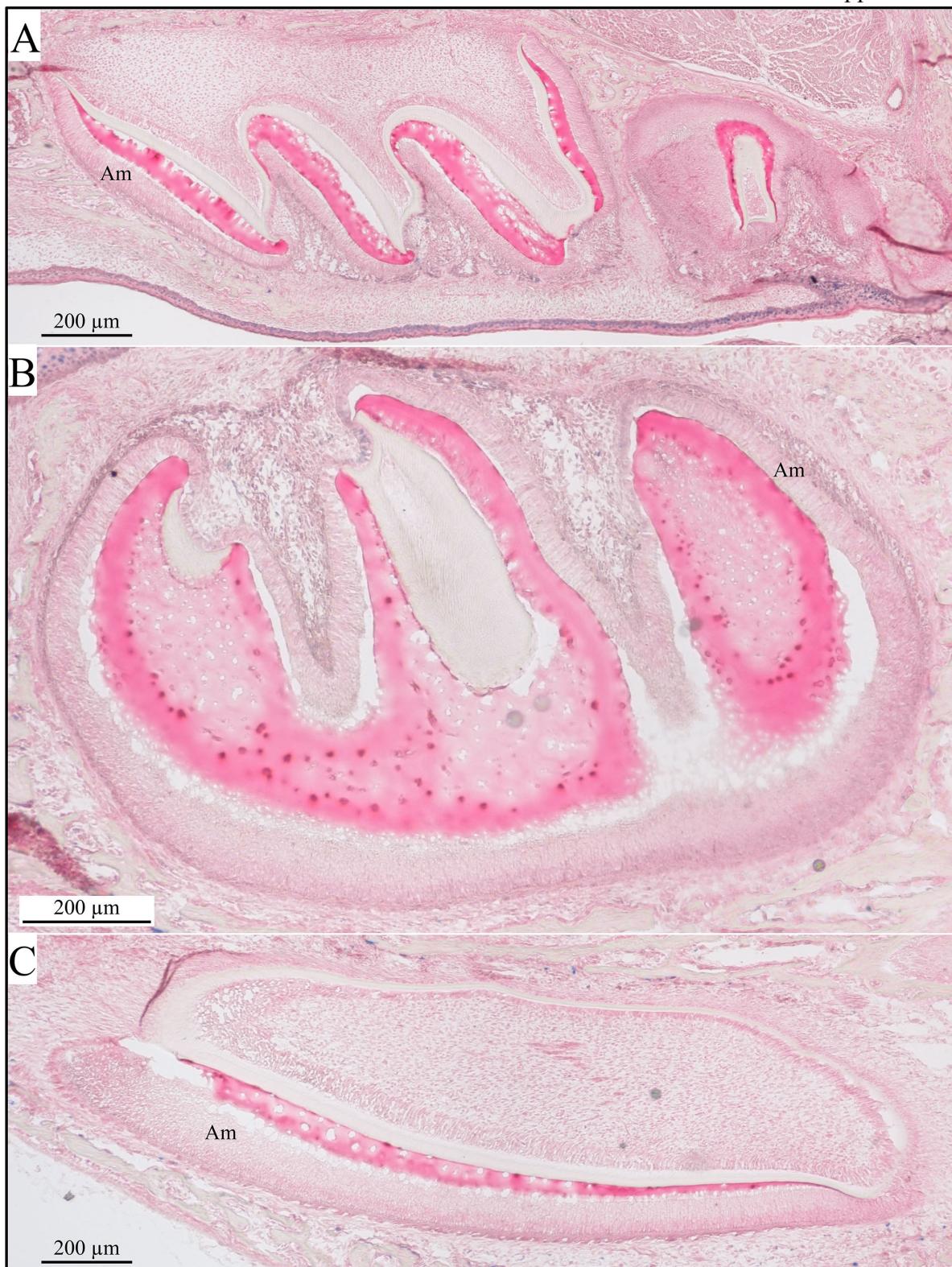
**Fig. S12.** *LacZ* Histochemistry of Developing PN5 *Fam83h<sup>+/−</sup>* Mouse Teeth. **A:** Low Magnification view of sagittal section of the head. The teeth are circled. **B:** Higher magnification of the mandibular incisor near cross-section. **C:** Higher magnification of the incisor cross-section. No X-gal histostaining is observed in the secretory stage ameloblasts (Am). **D:** PN5 *Fam83h<sup>+/−</sup>* mouse maxillary first and second molars. The secretory stage ameloblasts are negative except for the mesial cusp tip of the second molar. Note the positive (blue) staining in the nuclei of the oral mucosa.



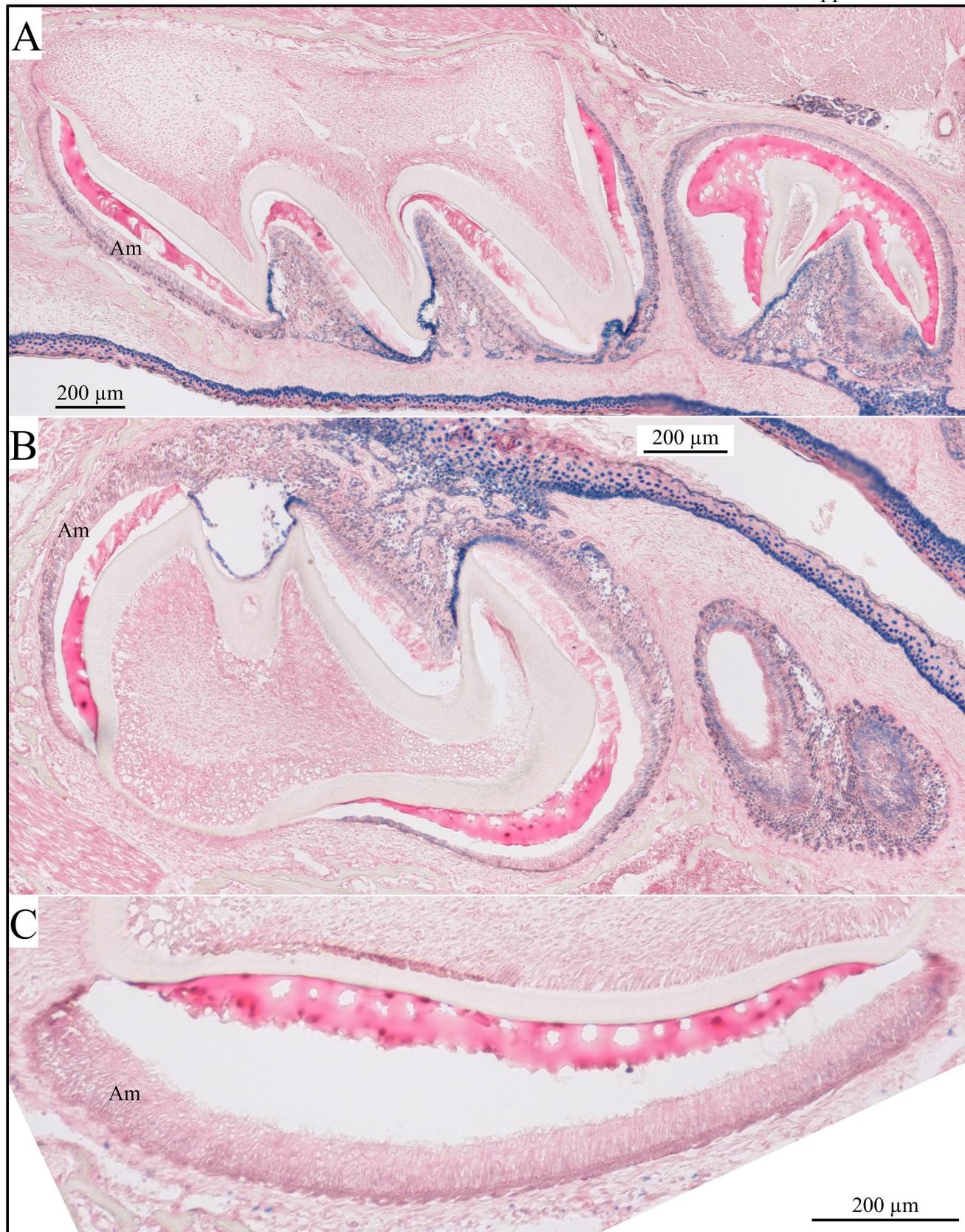
**Fig. S13.** *LacZ* Histochemistry of Developing PN5 *Fam83h* Null Mouse Teeth. **A:** Low Magnification view of sagittal section of the head. The teeth are circled. **B:** Higher magnification of the maxillary molars. Positive staining is observed in Ameloblasts on the mesial cusp of the 2<sup>nd</sup> molar. **C:** Higher magnification of the maxillary incisor. No X-gal histostaining is observed in the secretory stage ameloblasts (Am). **D:** Mandibular first molar showing only trace X-gal staining in Ameloblasts. Note the positive (blue) staining in the nuclei of the oral mucosa, which provides an internal positive control.



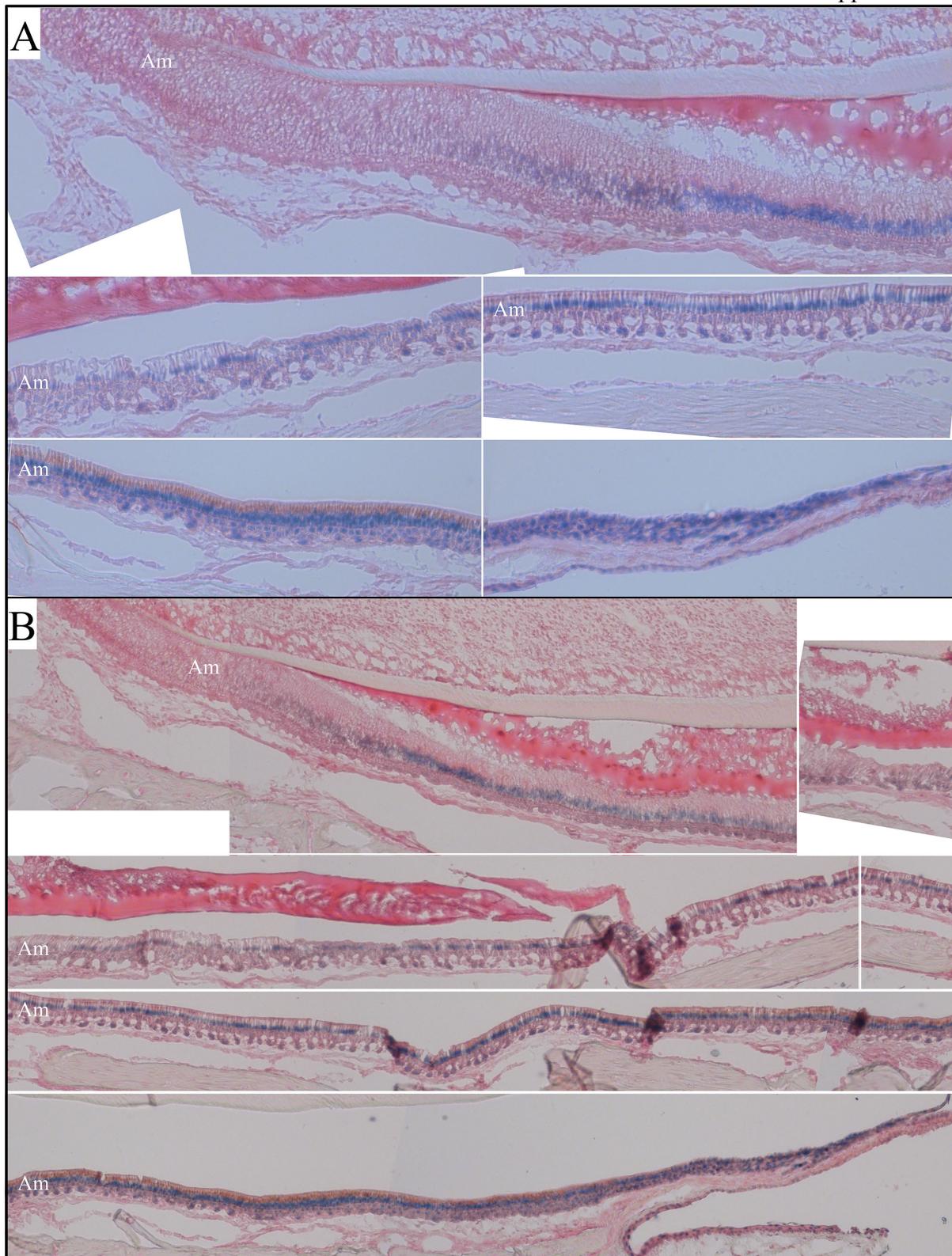
**Fig. S14.** LacZ Histochemistry of Developing PN6 *Fam83h* Null Mouse Teeth. **A:** Low Magnification view of the developing maxillary molars. The ameloblasts (Am) in these teeth are in the secretory stage of amelogenesis. No X-gal stain is observed in the maxillary first molar. Staining observed in ameloblasts at the distal cusp tip. Oral mucosa nuclei stain positive. **B:** Low magnification view of the developing mandibular molars. Most ameloblasts are negative except at the tip of the distal cusp in the first molar and the cusp slopes of the second molar and on the tip of the mesial cusp.



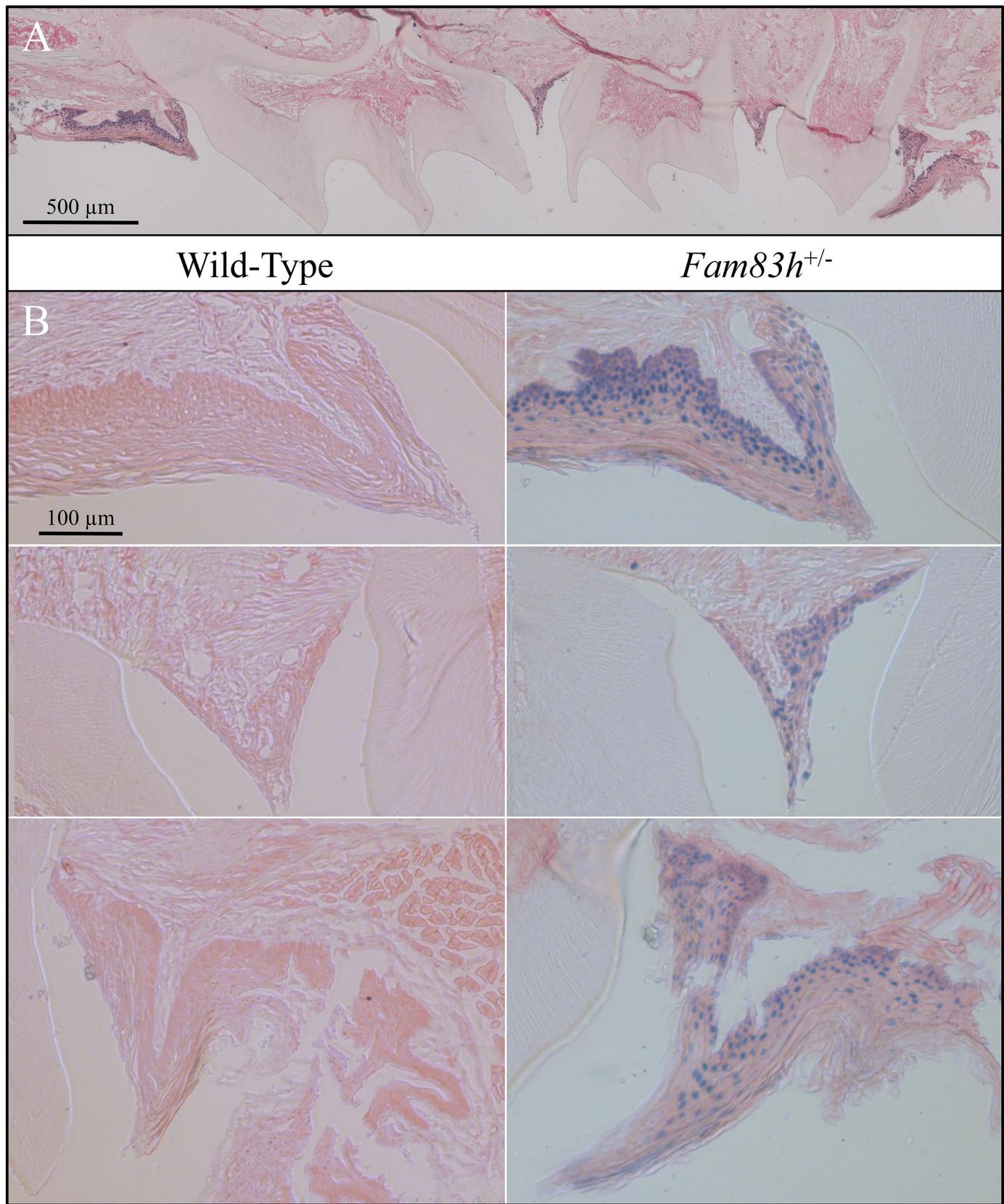
**Fig. S15.** *LacZ* Histochemistry of Developing PN9 *Fam83h* Null Mouse Teeth. **A-B:** Low Magnification views of the developing maxillary and mandibular first molars, which have ameloblasts (Am) in the maturation stage of amelogenesis. **C:** Low magnification view of the basal end developing mandibular incisor. No X-gal stain is observed in the maxillary first molar. Most ameloblasts are negative except at the tip of the distal cusp in the first molar and the cusp slopes of the second molar and on the tip of the mesial cusp.



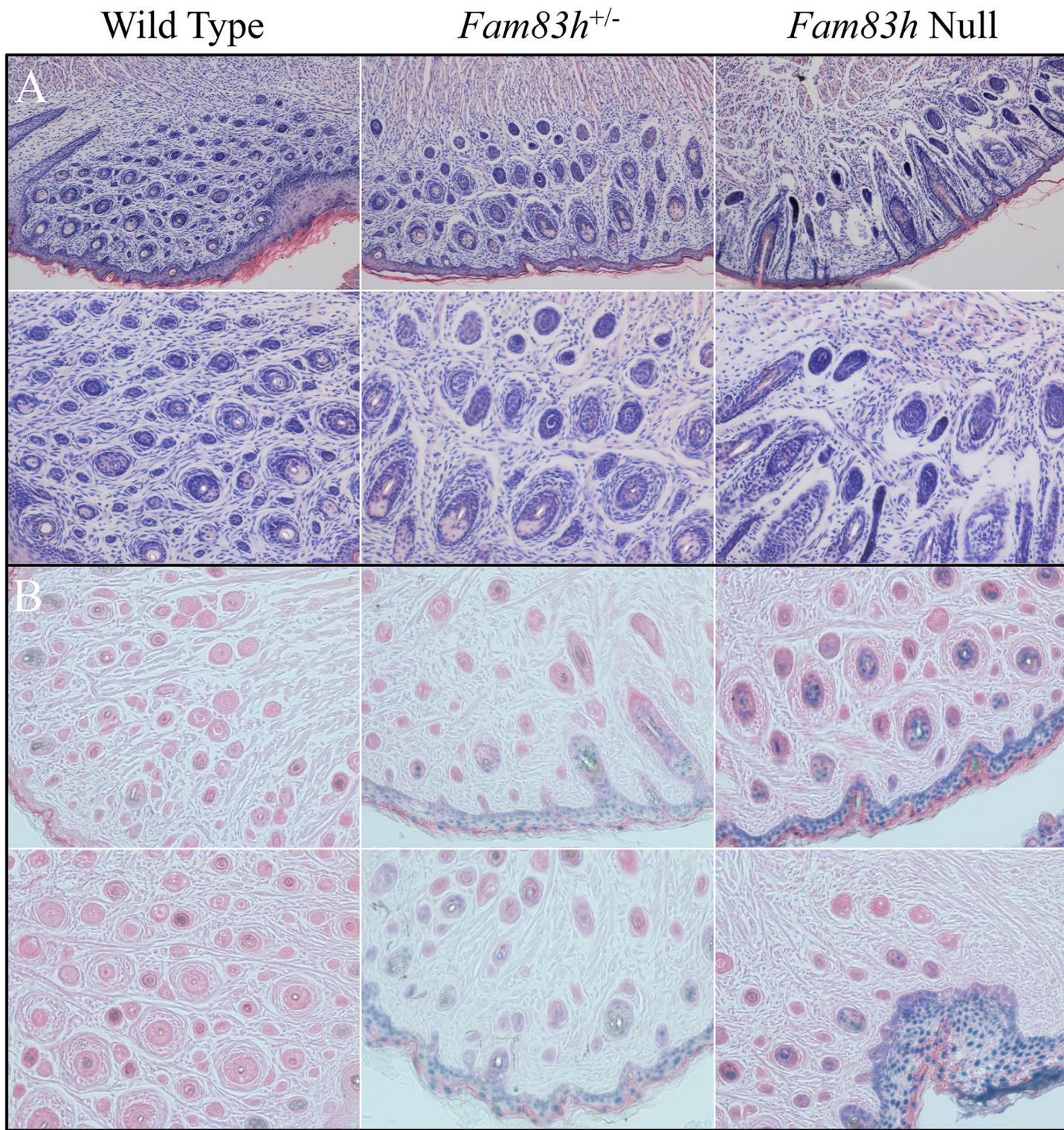
**Fig. S16.** *LacZ* Histochemistry of Developing PN11 *Fam83h* Null Mouse Teeth. **A-B:** Low Magnification views of the developing maxillary and mandibular first molars, which have ameloblasts (Am) in the maturation stage of amelogenesis. **C:** Low magnification view of the basal end developing mandibular incisor. No X-gal stain is observed in the maxillary first molar. Most ameloblasts are negative except at the tip of the distal cusp in the first molar and the cusp slopes of the second molar and on the tip of the mesial cusp.



**Fig. S17.** *LacZ* Histochemistry of 7-Week *Fam83h<sup>+/−</sup>* Mandibular Incisors. **A-B:** Low Magnification views of the developing maxillary and mandibular first molars, which have ameloblasts (Am) in the maturation stage of amelogenesis. **C:** Low magnification view of the basal end developing mandibular incisor. No X-gal stain is observed in the maxillary first molar. Most ameloblasts are negative except at the tip of the distal cusp in the first molar and the cusp slopes of the second molar and on the tip of the mesial cusp.



**Fig. S18.** *LacZ* Histochemistry of D28 *Fam83h*<sup>+/-</sup> Dental Papilla. **A:** X-gal stained D28 *Fam83h*<sup>+/-</sup> cryosection of the erupted maxillary molars. **B:** High magnification views X-gal stained cryosections from D28 wild-type and *Fam83h*<sup>+/-</sup> mice. The gingival epithelium stains positive.



**Fig. S19.** Histology and *LacZ* Histochemistry of PN5 Perioral Skin. **A:** H&E stained sections showed a decreasing number of vibrissae. The sebaceous glands look under-developed and disorganized. **B:** X-gal stained sections reported *Fam83h* expression in the cortex and root sheath areas of the vibrissae. All cell layers of the epidermis reported positive for *LacZ* stain, including the stratum spinosum and stratum basale.